



SPANISH ASSOCIATION OF PAEDIATRICS



Spanish consensus document on diagnosis, stabilisation and treatment of pediatric multisystem inflammatory syndrome related to SARS-CoV-2 (SIM-PedS)[☆]

Alberto García-Salido^{a,b,*}, Jordi Antón^{c,d}, José David Martínez-Pajares^{e,f}, Gemma Giralt García^{g,h}, Borja Gómez Cortés^{i,j}, Alfredo Tagarro^{k,l}, Grupo de trabajo de la Asociación Española de Pediatría para el Síndrome Inflamatorio Multisistémico Pediátrico vinculado a SARS-CoV-2

^a Sociedad Española de Cuidados Intensivos Pediátricos (SECIP)

^b Cuidados Intensivos Pediátricos, Hospital Infantil Universitario Niño Jesús, Madrid, Spain

^c Sociedad Española de Reumatología Pediátrica (SERPE)

^d Hospital Sant Joan de Déu, Institut de Recerca Sant Joan de Déu (IRSJD), Barcelona, Spain

^e Sociedad Española de Pediatría Hospitalaria (SEPHO)

^f Unidad de Pediatría, Hospital de Antequera, Área Sanitaria Norte de Málaga, Málaga, Spain

^g Sociedad Española de Cardiología Pediátrica y Cardiopatías Congénitas (SECPCC)

^h Hospital Universitario Hospital Vall d'Hebron, Hospital Universitario Vall d'Hebron, Barcelona, Spain

ⁱ Sociedad Española de Urgencias de Pediatría (SEUP)

^j Servicio de Urgencias de Pediatría. Hospital Universitario Cruces, Barakaldo

^k Sociedad Española de Infectología Pediátrica (SEIP)

^l Servicio de Pediatría, Hospital Universitario Infanta Sofía, Unidad Pediátrica de Investigación y Ensayos Clínicos (UPIC), Instituto de Investigación Sanitaria Hospital 12 de Octubre (IMAS12), Madrid, Spain

Received 12 August 2020; accepted 15 September 2020

Available online 14 January 2021

KEYWORDS

Multisystem inflammatory syndrome;
Paediatrics;
SARS-CoV-2

Abstract A new paediatric multisystem inflammatory syndrome, linked to SARS-CoV-2 (MIS-Paed), has been described. The clinical picture is variable and is associated with an active or recent infection due to SARS-CoV-2. A review of the existing literature by a multidisciplinary group of paediatric specialists is presented in this document. Later, they make recommendations on the stabilisation, diagnosis, and treatment of this syndrome.

© 2020 Asociación Española de Pediatría. Published by Elsevier España, S.L.U. This is an open access article under the CC BY-NC-ND license (<http://creativecommons.org/licenses/by-nc-nd/4.0/>).

[☆] Please cite this article as: García-Salido A, Antón J, David Martínez-Pajares J, García GG, Cortés BG, Tagarro A. Documento español de consenso sobre diagnóstico, estabilización y tratamiento del síndrome inflamatorio multisistémico pediátrico vinculado a SARS-CoV-2 (SIM-PedS). Estudio beenis. An Pediatr (Barc). 2021;94:118.

* Corresponding author.

E-mail address: citopenisis@yahoo.es (A. García-Salido).

PALABRAS CLAVE

Síndrome inflamatorio multisistémico;
Pediatría;
SARS-CoV-2

Documento español de consenso sobre diagnóstico, estabilización y tratamiento del síndrome inflamatorio multisistémico pediátrico vinculado a SARS-CoV-2 (SIM-PedS)

Resumen Se ha descrito un nuevo síndrome inflamatorio multisistémico pediátrico vinculado a SARS-CoV-2 (SIM-PedS). Este cuadro presenta una expresividad clínica variable y se asocia a infección activa o reciente por SARS-CoV-2. En este documento se revisa la literatura existente por parte de un grupo multidisciplinar de especialistas pediátricos. Posteriormente se realizan recomendaciones sobre estabilización, diagnóstico y tratamiento de este síndrome.

© 2020 Asociación Española de Pediatría. Publicado por Elsevier España, S.L.U. Este es un artículo Open Access bajo la licencia CC BY-NC-ND (<http://creativecommons.org/licenses/by-nc-nd/4.0/>).

Introduction

It is known infection by the novel coronavirus (SARS-CoV-2) in the paediatric population is usually mild.^{1,2} In Spain, paediatric patients aged less than 15 years account for 0.4% of hospital admissions, 0.7% of intensive care unit admissions and 0.15% of deaths to date.¹ However, since the beginning of May 2020, a small number of paediatric patients have developed a hyperinflammatory syndrome with a variable presentation.³ This syndrome is characterised by clinical and laboratory features similar to those found in Kawasaki disease (KD), toxic shock syndrome (TSS) or macrophage activation syndrome (MAS).^{4–7} Official health agencies, such as the Centers of Disease Prevention and Control (CDC) of the United States, the World Health Organization or the Royal College of Paediatrics and Child Health (RCPCH) have defined its general characteristics.^{5–7}

This novel syndrome seems to be associated with active or recent infection by SARS-CoV-2. Most affected patients have positive IgG antibody tests and elevated inflammatory markers, which suggests immune dysregulation as opposed to a direct pathogenic effect of the virus.^{6,7} In this document, we will use the term paediatric multisystem inflammatory syndrome – temporally associated with SARS-CoV-2 (PIMS-TS) to refer to this condition, also known widely as multisystem inflammatory syndrome in children (MIS-C).

Definitions and differential diagnosis

The definition of PIMS-TS differs slightly between these public health agencies (Table 1).^{4–6} This diagnosis should be contemplated in areas with a current or recent high incidence of cases of SARS-CoV-2 or coronavirus disease 2019 (COVID-19).⁸ It has significant overlap with the different clinical forms of KD (complete KD, incomplete KD, KD shock syndrome [KDSS]). Therefore, some experts recommend considering PIMS-TS in patients meeting the diagnostic criteria for any of the forms of KD.⁹

The following features are particularly relevant for suspicion of PIMS-TS: diagnostic criteria for complete or incomplete KD at any age, presence of gastrointestinal manifestations (vomiting, nausea, abdominal pain, diarrhoea), elevation of acute phase reactants, shock, hypotension,

myocardial dysfunction, lymphopenia, anaemia and thrombocytopenia (Table 2).¹⁰

The differential diagnosis should always include the following:

- Bacterial sepsis.
- Other viral infections (adenovirus, enterovirus, measles in unvaccinated individuals).
- Acute abdomen.
- Streptococcal or staphylococcal TSS.
- Myocarditis caused by other microorganisms.
- KD unrelated to SARS-CoV-2.
- Drug hypersensitivity reaction (Stevens-Johnson syndrome).
- Other systemic rheumatic disorders (systemic onset juvenile idiopathic arthritis and other autoinflammatory or autoimmune diseases).
- Primary or secondary haemophagocytic lymphohistiocytosis (HLH).

Initial stabilization and care

Initial care and stabilization will follow the ABCDE approach. The airway is usually patent, unless the patient presents with altered level of consciousness. Administer supplemental oxygen as needed. Monitor the oxygen saturation (SatO₂), along with the expired carbon dioxide (EtCO₂) when possible. Prepare the equipment and drugs needed for rapid sequence intubation.

Monitor the respiratory rate (RR). Tachypnoea without breathing difficulty is a possible feature in these patients (as compensation of the metabolic acidosis associated with shock). Rule out lung infection or cardiogenic pulmonary oedema. Measure and document the blood pressure (BP), heart rate (HR) and peripheral perfusion (capillary refill time, skin temperature and colour, perfusion index). Establish peripheral vascular access (ideally, 2 lines). In case it cannot be establish, consider a peripheral catheter (ideally, 2). If this fails, consider placement of an intraosseous catheter. Fig. 1 presents the recommended algorithm for management of haemodynamic support. The level of consciousness, pupillary reflex, glycaemia and severity of pain must be assessed in every patient. It is also necessary to

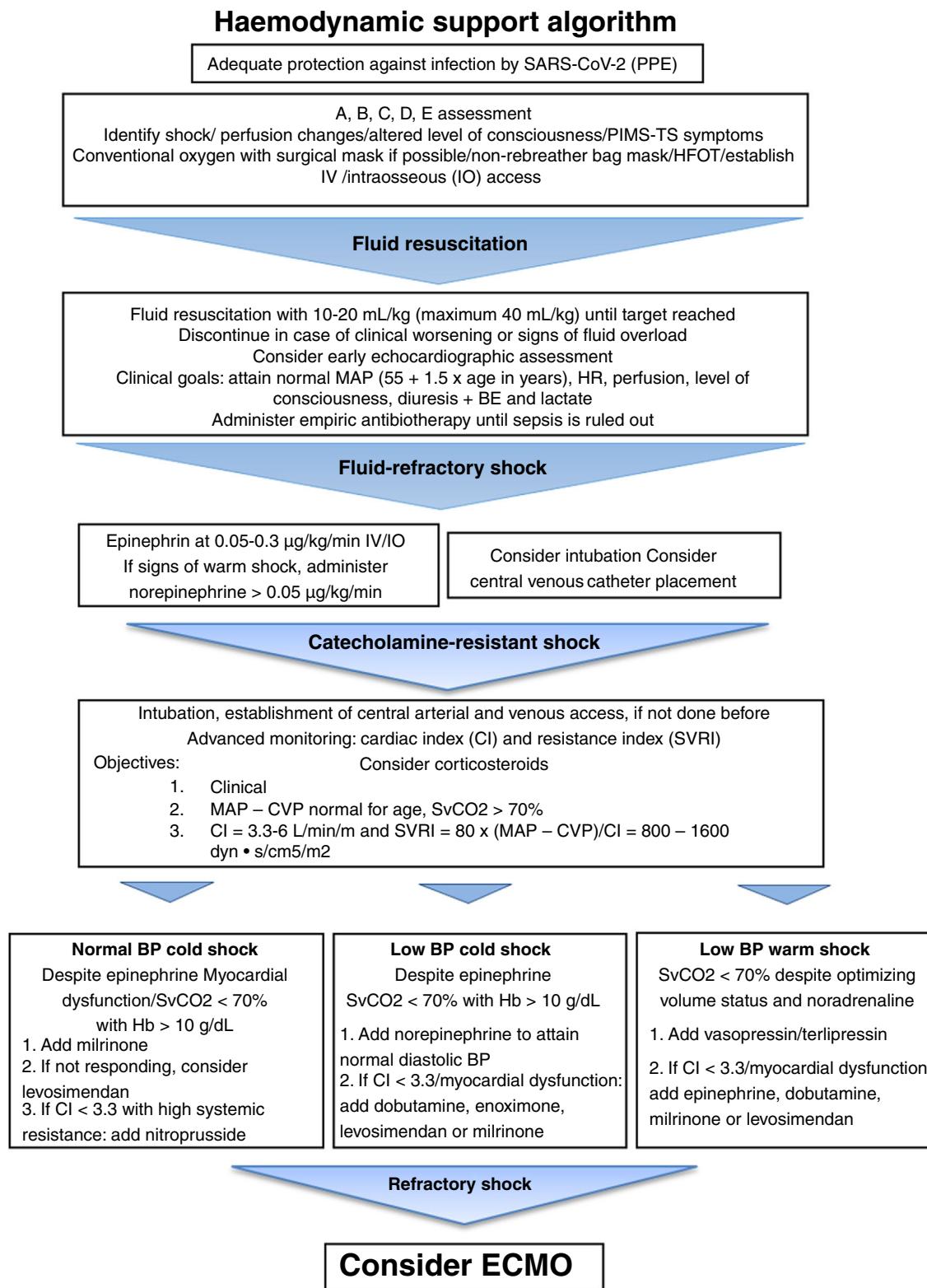


Fig. 1 Algorithm for haemodynamic support in patients with PIMS-TS. BE, base excess; CI, cardiac index; CVP, central venous pressure; ECMO, extracorporeal membrane oxygenation; HFOT, high-flow oxygen therapy; HR, heart rate; IV, intravenous; MAP, mean arterial pressure; PPE, personal protective equipment; RR, respiratory rate; SBP, systolic blood pressure; $SvCO_2$, central venous oxygen saturation; SVRI, systemic vascular resistance index.

Adapted from the American College of Critical Care Medicine Clinical Practice Parameters for Hemodynamic Support of Pediatric and Neonatal Septic Shock. Critical Care Medicine 2017;45:1062-1093.

Table 1 Case definition of PIMS-TS of the World Health Organization (WHO), Centres for Disease Control and Prevention (CDC) and Royal College of Paediatrics and Child Health (RCPCH).

WHO	Patient aged \leq 19 years with fever \geq 3 days AND at least 2 of the following criteria: Rash, bilateral nonpurulent conjunctivitis, or mucocutaneous inflammation signs (oral, hands, or feet) Hypotension or shock Cardiac dysfunction, pericarditis, valvulitis, or coronary abnormalities (including echocardiographic findings or elevated troponin/BNP) Evidence of coagulopathy (prolonged PT or PTT; elevated D-dimer) Acute gastrointestinal symptoms (diarrhoea, vomiting, or abdominal pain) AND elevated markers of inflammation (eg, ESR, CRP, or procalcitonin) AND no other obvious microbial cause of inflammation, including bacterial sepsis and staphylococcal/streptococcal toxic shock syndromes AND Evidence of SARS-CoV-2 infection (positive SARS-CoV-2 RT-PCR, serology, antigen test) or contact with an individual with COVID-19
CDC	Patient aged $<$ 21 with fever $>$ 24 h AND evidence of severe disease requiring hospital admission with involvement of more than 2 organs or systems (cardiovascular, respiratory, haematologic, gastrointestinal, dermatologic or neurologic) laboratory evidence of inflammation (elevated CPR, ESR, fibrinogen, PCT, D-dimer, ferritin, LDH or IL-6, neutrophilia, lymphopenia or hypoalbuminaemia) AND lack of alternative plausible diagnosis AND evidence of recent or current SARS-CoV-2 infection or exposure (positive SARS-CoV-2 RT-PCR, serology, antigen test) or of COVID-19 exposure within the 4 weeks prior to the onset of symptoms
RCPCH, UK	Persistent fever AND signs of inflammation (neutrophilia, elevated CRP and lymphopenia) AND evidence of single or multiple organ dysfunction (shock, cardiac, respiratory, renal, gastrointestinal or neurologic disorders), with additional features (see table in the case definition). It may include children fulfilling full or partial criteria for Kawasaki disease Exclusion of any other microbial cause, including bacterial sepsis, staphylococcal or streptococcal shock syndromes, infections associated with myocarditis such as enterovirus SARS-CoV-2 PCR testing may be positive or negative

BNP, B-type natriuretic peptide; COVID, coronavirus disease; CRP, C-reactive protein; ESR, erythrocyte sedimentation rate; IL-6, interleukin 6; LDH, lactate dehydrogenase; PCT, procalcitonin; PPT, partial prothrombin time; PT, prothrombin time; RT-PCR, reverse transcription-polymerase chain reaction.

Table 2 Most frequent clinical manifestations and laboratory findings.

Clinical manifestations
Fever (in nearly 100% of cases); fever $>$ 3 days (a shorter duration does not rule out PIMS-TS)
Gastrointestinal symptoms (> 50%): abdominal pain, vomiting, diarrhoea
Rash (scarlatiniform rash, erythroderma, erythema multiforme, livedo reticularis), non-exudative conjunctivitis, mucosal abnormalities, peripheral abnormalities (>2/3 patients)
Shock, tachycardia, hypotension, hypoperfusion (approximately half of patients)
Headache, meningism, confusion (10–20%)
Respiratory symptoms: cough, dyspnoea (30-60%)
Laboratory findings
Complete blood count: leucocytosis with lymphopenia, neutrophilia and thrombocytopenia
Inflammatory markers: elevation of CRP, ESR, ferritin, fibrinogen, LDH, IL-6. Normal or elevated PCT normal (in absence of bacterial infection)
Coagulation: fibrinogen, D-dimer elevation
Blood chemistry: hyponatremia, hypoalbuminemia, transaminase elevation (ALT, AST)
Cardiac markers: high elevation of NT-proBNP (> 200 ng/L), elevation of cardiac enzymes (troponin-I, CK-MB)

ALT, alanine aminotransferase; AST, aspartate transaminase; CK-MB: creatine kinase myocardial band; CPR, C-reactive protein; ESR, erythrocyte sedimentation rate; IL-6, interleukin 6; LDH, lactate dehydrogenase; NT-proBNP, N-terminal pro-B-type natriuretic peptide; PCT, procalcitonin; PPT, partial prothrombin time; PT, prothrombin time.

Table 3 Recommended diagnostic tests and most frequent abnormalities.

Test	Abnormalities
<i>Complete blood count</i>	Leucocytosis (usually < 20 000/mm ³) with lymphopenia Anaemia variable
<i>Electrolyte panel</i>	Mild thrombocytopenia (normally > 50 000/mm ³)
<i>Liver panel and albumin</i>	Hyponatraemia Transaminase elevation Hypoalbuminemia
<i>Cardiac markers^a</i>	Elevation of pro-BNP (> 35 pg/mL) or NT-proBNP (> 125 pg/mL) and ultrasensitive troponin (> 14 ng/L)
<i>Blood gases</i>	Metabolic/respiratory acidosis, depending on clinical condition
<i>Inflammatory markers^a</i>	Elevated CPR (> 20 mg/L), PCT (> 0.5 µg/mL), IL-6 (> 8.5 pg/mL) and ferritin (> 120 mg/dL)
<i>Coagulation study</i>	Increased fibrinogen (> 400 mg/dL) Significant D-dimer elevation D (> 500 ng/mL) Abnormal PT/PPT
<i>Blood culture</i>	Usually negative
<i>Other</i>	Coinfections
[0,1–2] Urine culture	
[0,1–2] PCR respiratory panel in nasal aspirate sample	
[0,1–2] Stool culture/PCR for detection of pathogens in stool	

CPR, C-reactive protein; IL-6, interleukin 6; PCT, procalcitonin; PPT, partial prothrombin time; PT, prothrombin time.

^a Suggested values, consider the reference values used at the hospital.

assess for the presence of exanthema and petechiae and measure the temperature.¹¹

Any patient with suspected PIMS-TS should be transferred to a hospital allowing multidisciplinary management. In severe cases, transfer to a hospital with a paediatric intensive care unit (PICU) is recommended.^{4–6,12}

Diagnostic tests unrelated to SARS-CoV-2

Alternative infectious agents that could be causing the disease must be ruled out. Table 3 details the recommended diagnostic tests and the most frequent abnormalities found in patients with suspected PIMS-TS.^{10–13} Patients with shock may present with lymphopenia, anaemia and thrombocytopenia and increased levels of ferritin, LDH and D-dimer D (DD).¹⁰ Patients with coronary aneurysms may present with even lower white blood cell and lymphocyte counts associated with elevation of C-reactive protein (CPR).^{10–13,14}

Compared to KD or TSS, PIMS-TS usually presents with a greater elevation of inflammatory markers.¹⁰ Specifically, compared to complete KD, PIMS-TS is associated with higher levels of CRP, ferritin, inflammatory cytokines and N-terminal pro b-type natriuretic peptide (NT-proBNP) and a higher frequency of lymphopenia and thrombocytopenia, with no differences in DD levels.^{14–17}

Another frequent feature is an increased neutrophil-lymphocyte ratio. There is a decrease in the CD4+, CD8+ and natural killer (NK) T cell counts. This phenomenon could be associated with the infiltration of the extracellular space by these innate immune system lymphocytes.¹⁸ When it comes to the cytokines, PIMS-TS manifests with elevation of interleukin (IL)-1 in its soluble form (IL-1 β) and IL-6.^{19–21}

Diagnostic tests related to SARS-CoV-2

Most patients with PIMS-TS have positive results in one or more diagnostic tests for detection of acute or past infection by SARS-CoV-2. In every patient with suspected PIMS-TS, at least 1 respiratory sample should be obtained to perform a reverse-transcriptase polymerase chain reaction (RT-PCR) test. It is widely accepted that the samples that offer the highest sensitivity for detection of SARS-CoV-2 infection are bronchoalveolar lavage or endotracheal aspirate samples. Nasal swab, throat swab or nasal wash samples are also acceptable.^{22,23} If the RT-PCR test is negative and the clinical suspicion is high, we recommend repetition of the test in the next 24–48 hours. In case of clinical worsening requiring invasive mechanical ventilation, we recommend collection of an endotracheal aspirate sample.²²

We also recommend serologic testing of all patients, independently of RT-PCR test performance or results. The yield of serology is greater after a minimum of 10 days from the onset of symptoms. In patients with negative results of both RT-PCR and serology with a high clinical suspicion, we recommend repetition of serologic testing 3–4 weeks after admission. Previous studies have reported that 26 to 55% of patients with PIMS-TS have positive RT-PCR tests and up to 90% have positive for IgG antibody tests.^{10–17} Antibodies can be detected from 10 to 15 days after infection, and cumulative seroconversion peaks at about 16–21 days.^{22,23}

Imaging tests

Performance of plain chest radiograph or a lung ultrasound scan is indicated in patients with respiratory symptoms or to assess placement of indwelling devices (endotracheal tube or central lines).²⁴ The chest radiograph can detect

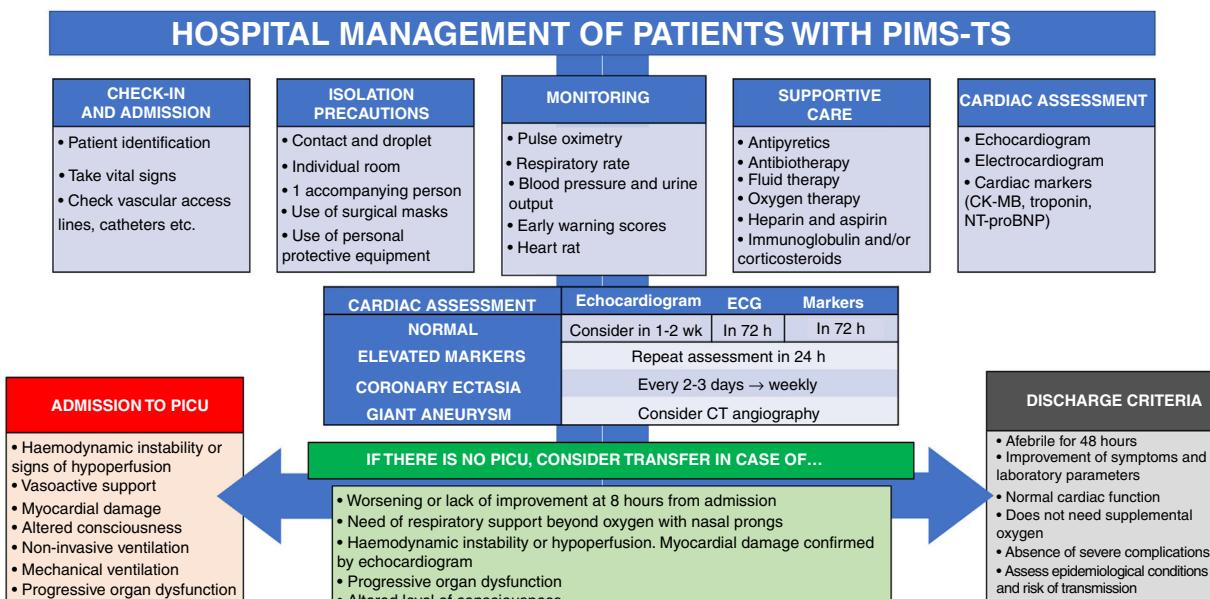


Fig. 2 Hospital management and treatment of PIMS-TS.

abnormalities in a high percentage of cases.⁸⁻²⁵ Some of the radiographic features reported in the literature included pulmonary infiltration, patchy consolidation, ground glass opacities or interstitial changes, atelectasis and pleural effusion. The lung ultrasound scan can detect patterns compatible with pneumonia or a buildup of fluid in the lungs. Routine performance of a thoracic computed tomography (CT) scan is not recommended, except in patients with complicated pneumonia, significant worsening or who are immunocompromised.

An abdominal ultrasound or CT scan is indicated in patients with manifestations suggestive of acute abdomen to detect potential complications and assess the need of surgery. Ileocolitis has been described in some patients with PIMS-TS.

Cardiac function tests

We recommend performance of an echocardiographic study in all patients with PIMS-TS. Echocardiographic abnormalities are not usually found in patients with mild forms. The abnormalities described in patients with more severe disease include left ventricular systolic dysfunction, right ventricular dysfunction, mitral valve insufficiency, pericardial effusion and coronary artery (CA) dilatations or aneurysms.¹³ The assessment of the CAs includes calculation of CA size z-scores and classification based on the National Consensus Document on Kawasaki Disease.²⁶ A high proportion of patients exhibit elevation of troponin I or T (55-68%) and BNP/NT-proBNP (83-100%), with significantly higher values compared to patients with shock.^{3,10-19}

The electrocardiographic features are generally non-specific. Patients with PIMS-TS may exhibit abnormalities suggestive of myocardial involvement, such as low voltage, ST segment abnormalities and prolongation of the T wave or the QT interval. Different degrees of atrioventricular block and supraventricular and ventricular arrhythmias have also

been reported.⁹⁻¹³ Cardiovascular magnetic resonance imaging (MRI) is not indicated during the acute phase of disease. Performance of MRI should be contemplated based on the suspicion of cardiac involvement, patient safety during the test and availability of MRI in the hospital.

Hospital admission and care

All patients with suspected PIMS-TS should be admitted to the ward for observation and treatment, if applicable. All efforts must be made to keep the staff that enters in contact with the patient to the minimum necessary. The staff in contact with a patient with PIMS-TS must wear, at minimum, a FFP2 mask, disposable gowns, gloves and eye protection until the patient has at least 2 negative RT-PCR test results. A high percentage of patients will have negative RT-PCR results. This is indicative of a low or no risk of transmission. In patients with potential active infection despite a negative RT-PCR test, a second RT-PCR test must be done. Fig. 2 summarizes the recommendations on isolation measures, monitoring and treatment.

Critical patients

If admission to PICU is necessary, the patient should be placed in an isolation room, preferably one with negative pressure.²⁷ If performance of aerosol-generating procedures is necessary, the care staff will wear FFP3 masks, a full-face powered-air-purifying respirator, eye goggles, disposable coveralls or caps, a water-resistant disposable gown or, if the gown is not water-resistant, a water-resistant apron, and gloves. If intubation is required, use double gloves.

The patient will be accompanied by one family member or caregiver that will adhere to the isolation and hygiene measures as directed by staff and will wear, at minimum,

a surgical mask, gown and gloves. If the RT-PCR is negative, modification of the required protective measures will be considered, always maintaining contact and droplet precautions.

Patients admitted to the PICU will be monitored per protocol according to their condition and disease severity. In severe cases, consider early establishment of central arterial and venous access.²⁸

Respiratory support

Respiratory distress occurs in 70% of cases, and the need of respiratory support is usually associated with cardiovascular involvement or haemodynamic instability.²⁴ Administer supplemental oxygen through nasal prongs under a surgical mask to maintain the SatO₂ in the 94 to 98% range. If this is not sufficient due to the condition of the patient or the arterial blood gas values, consider high-flow oxygen therapy (HFOT) or non-invasive ventilation (NIV). Respiratory support must always be provided while maintaining isolation and protective measures.

In case of hypoxaemia in absence of hypercapnia, consider initiation of HFOT.

If HFOT is ineffective or as an alternative option, initiate NIV with continuous positive airway pressure (CPAP) with an oronasal mask, full-face mask or helmet (the latter provides the best seal). If the patient also has hypercapnia, consider bilevel positive airway pressure (BIPAP).²⁹

If there is no clear improvement in clinical parameters (HR, RR, respiratory distress) and oxygenation (PaO₂/FiO₂, SatO₂/FiO₂) within a few hours, we recommend early intubation. Early intubation should also be considered in case of altered level of consciousness or fluid-refractory or catecholamine-resistant shock.²⁹ We recommend the following initial settings for respiratory support: tidal volume of 4 to 8 mL/kg, identification of the optimal positive end-expiratory pressure to achieve adequate alveolar recruitment, plateau pressures of less than 30 cmH₂O and a driving pressure of less than 15 cmH₂O.²² If the patient develops moderate-to-severe acute respiratory distress, follow the recommendations for lung protection with permissive hypercapnia, use of the prone position and neuromuscular blockade. Nitric oxide is reserved for patients with refractory hypoxaemia, especially if it is associated with pulmonary hypertension.³⁰

Haemodynamic support

Haemodynamic instability is usually associated with vaso-plegic syndrome or heart failure.^{5,31} Fig. 1 summarises the recommendations for haemodynamic support and its goals.¹¹ In patients with heart failure administer epinephrine and consider addition of milrinone or, in case of moderate to severe heart failure, levosimendan.

Pharmacotherapy

Immunomodulatory therapy will be given with a stepwise approach, starting with intravenous immunoglobulin (IVIG)

or corticosteroids as first-line treatment.^{24–32} In severe or refractory cases, we recommend combining both.

Immunomodulatory therapy

Intravenous immunoglobulin

Intravenous immunoglobulin should be given at a dose of 2 g/kg, especially in patients meeting the criteria for KD or TSS.^{12–17} Haemodynamic instability can be treated with 1 g/kg/day for 2 days. If the fever persists, consider administration of a second dose 36 h after the first.^{17–31}

Systemic corticosteroid therapy

Intravenous corticosteroid therapy is indicated both as the first step of treatment and in patients that do not respond to the initial dose of IVIG. Early administration should be considered in patients with risk factors for development of coronary aneurysms or laboratory results compatible with MAS.^{20–26}

- Mild-moderate disease: intravenous methylprednisolone at 1–2 mg/kg/day for 3–5 days followed by discontinuation of methylprednisolone. In patients requiring treatment for 6 or more days or with persistent symptoms or elevation of inflammatory markers, step up to oral prednisone with tapering off over 2–3 weeks.
- Severe disease (shock, especially in patients requiring high doses of inotropes/vasopressors): intravenous methylprednisolone at 1–2 mg/kg/day for 3–5 days or intravenous methylprednisolone at 30 mg/kg/day for 1–3 days (maximum of 1 g). If the patient responds well, continue treatment with oral prednisone at 1–2 mg/kg/day with tapering of the dose until the inflammatory markers normalize.
- Kawasaki-like disease: combine corticosteroids and IVIG in patients at high risk of IVIG resistance (male, age < 12 months, CPR > 100 mg/L, platelet count < 300 000/mm³, alanine aminotransferase > 100 IU/L, neutrophil count > 80%, sodium < 133 mmol/L).⁹

Interleukin-1 inhibitors

An IL-1 receptor antagonist (anakinra) has been used successfully in patients with severe pneumonia and hyperinflammation associated with SARS-CoV-2 infection,³³ MAS and KD refractory to IVIG/corticosteroid therapy. Thus, it may be beneficial to patients with PIMS-TS. It is considered a safe drug on account of its short half-life, quick action and infrequent association with bacterial superinfections. Although it is distributed in syringes for subcutaneous administration, it has been delivered intravenously in severely ill patients.^{5–10} The duration of treatment would be of 5–14 days depending on the clinical response (Table 4).

Tumour necrosis factor alpha inhibitors

Infliximab has been proposed as an alternative treatment in patients with IVIG-resistant KD, but in clinical trials, while it achieved quicker improvement of symptoms and laboratory parameters, it did not improve long-term cardiovascular

Table 4 Dosage, precautions and route of administration of immunomodulator drugs.

	Drug	Dose	Dilution	Adverse effects	Precautions
Anti- IL-1	Anakinra	Subcutaneous 2 mg/kg/day with progressive increase to 8 mg/kg/day every 12 h (maximum of 400 mg/day) Intravenous 2 options: 1) similar to subcutaneous 2) continuous infusion < 20 kg: single 2 mg/kg dose, followed by 0.02 mL/kg/h > 20 kg: single 2 mg/kg dose, followed by 0.01 mL/kg/h Maximum dose 400 mg/day	If intravenous, dilute with PS to concentration of 4–36 mg/mL	Local reaction at injection site, flu-like illness, neutropenia, headache, myalgia, higher vulnerability to infection	Local cooling at injection site. Monitor transaminase levels
Anti-TNF- α	Infliximab	5 mg/kg	Dilute the reconstituted dose in 250 mL of normal saline. Administer over 2 h	Anaphylaxis, infection	Consider premedication with antihistamine and corticosteroid to prevent infusion reaction
Anti- IL-6	Tocilizumab	Single dose < 30 kg: 12 mg/kg > 30 kg: 8 mg/kg (maximum of 800 mg)	< 30 kg: dilute in 50 mL of normal saline > 30 kg: dilute in 100 mL of normal saline Administer over 1 h	Neutropenia, thrombocytopenia, hypertransaminaemia, infections, intestinal perforation	Close monitoring of concurrent infections (prevent CPR elevation) Do not use if platelets < 100 000, neutrophils < 500 or AST/ALT > 3 times the baseline

^aIf the dose of anakinra is > 100 mg/day, administer every 8–12 h or as continuous infusion.

^bThe intravenous route is preferred for doses > 100 mg/day or in patients with a platelet count < 20 000, haemorrhagic complications or severe oedema.

outcomes.^{10,11} There are few data on its use in patients with PIMS-TS (**Table 4**).

Interleukin-6 inhibitors

Tocilizumab is approved for treatment of CAR T-cell-associated cytokine release syndrome and several trials are currently underway of its use in patients with SARS-CoV-2 pneumonia. It has been used in select cases of PIMS-TS, but the use of tocilizumab in patients with KD could accelerate the development of coronary aneurysms^{34,35} (**Table 4**).

Antiviral therapy

At the time of this writing, there is no evidence on the efficacy or safety of any antiviral used for treatment of SARS-CoV-2 infection in children. We recommend against the use of hydroxychloroquine^{36,37} or hydroxychloroquine combined with azitromycin.^{37,38} While the role of SARS-CoV-2 in the development of PIMS-TS remains unclear, treatment with remdesivir should be contemplated in case of active infection, high suspicion of SARS-CoV-2 or severe disease

(**Table 5**). Remdesivir can be administered to paediatric patients in the context of a clinical trial or after obtaining authorization for expanded access.³⁹

Antithrombotic and antiaggregant therapy

Indications for low molecular weight heparin

Consider prophylaxis with low molecular weight heparin in the following cases³⁹:

- a) DD \geq 6 times the upper limit of normal.
- b) Immobilized patient.
- c) Presence of giant aneurysms.
- d) Severe left ventricular dysfunction (ejection fraction < 30%).
- e) Personal or family history of thromboembolism.
- f) Personal history of arterial ischaemia (peripheral, coronary or cerebrovascular).

Administer subcutaneous enoxaparin at 1 mg/kg/day (in case of renal insufficiency with a glomerular filtration rate < 30 mL/min/m², dose of 0.25 mg/kg/12 h). Measure anti-factor Xa levels at 48–72 h (recommended range,

Table 5 Dosage and adverse effects of antiviral therapy.

Drug	Indication	Dose	Adverse effects	Monitoring
Remdesivir (intra-venous)	Clinical trial/ compassionate use	Weight: 2.5-40 kg Loading dose: 5 mg/kg/24 h (1 bolus) Maintenance dose: 2.5 mg/kg/24 h Weight \geq 40 kg Loading dose: 200 mg/24 h (1 bolus) Maintenance dose: 100 mg/24 h	Hypertransaminasaemia Altered renal function	Transaminases Renal function

0.3–0.49). Continue until symptoms resolve and DD levels have normalized based on the reference values used at the specific hospital.

In patients with thromboembolism or suspected deep vein thrombosis, administer enoxaparin at a dose of 1 mg/kg every 12 h delivered subcutaneously. When given in anti-coagulant doses (0.5–1 anti-Xa), measure anti-Xa at 48 h (adjust dose based on reference value). Maintain throughout the hospital stay and consult with haematology department prior to discharge.

Acetylsalicylic acid

There are 2 possible indications:

- **Anti-inflammatory:** patients with PIMS-TS meeting the criteria for complete or incomplete KD. Prescribe along with IVIG. Start treatment with oral acetylsalicylic acid (ASA) at a dose of 30–50 mg/kg/day given at 6-h intervals until the patient is afebrile for 48 h. At this time, reduce to a dose appropriate for antiaggregant therapy of 3–5 mg/kg/day, also by the oral route. Maintain this dose until 6–8 weeks from the onset of symptoms and after verifying normalization of the platelet count, levels of acute phase reactants and echocardiographic parameters.²⁶
- **Antiaggregant:** in patients with PIMS-TS that are severely ill, have aneurysms, with clinical or laboratory findings indicative of inflammation or thrombocytosis with a platelet count greater than 700 000/mm³, consider administration of ASA at an antiaggregant dose for 6 weeks (confirm normalization of echocardiographic findings at the end of this period). This recommendation is based on the description of coronary abnormalities in patients that do not meet the criteria for complete or incomplete KD.¹⁰

Rescue therapy with ECMO

Patients with PIMS-TS may be eligible for extracorporeal membrane oxygenation if they fail to respond to maximal conventional therapies, their condition is deemed reversible and they have no absolute contraindications for ECMO.⁴⁰ These patients must be referred before their condition precludes conventional transport. When the latter occurs, consider transport under ECMO with cannulation at the referring hospital performed by qualified staff.

Discharge and followup

Assess the potential risk of transmission of SARS-CoV-2 in the household. Patients with a negative RT-PCR test and a positive IgG test are not considered infectious. Also, positive detection of IgG antibodies combined with a positive RT-PCR test or seroconversion after negative PCR results is currently interpreted as detection of dead viral remnants that are not infectious. We recommend followup by a multidisciplinary team at the outpatient level, with involvement of the primary care paediatrician (Fig. 2).

Conflicts of interest

Jordi Antón has received research grants, speaker and consultant fees and financial support to attend medical congresses from Sobi and Roche. Esmeralda Núñez has participated in educational activities sponsored by AbbVie, Roche and Sobi. Inmaculada Calvo has participated in educational activities sponsored by Novartis, AbbVie, Sobi, Roche and GSK and collaborated in workshops offered by GSK, Novartis and Sobi. Javier Pérez-Lescure Picarzo has received fees from MSD for delivery of educational programmes. The rest of the authors have no conflicts of interest to declare.

Acknowledgments

We thank all health care professionals involved in paediatric care, patients and their families.

Appendix A. Members of the Working Group on Paediatric Multisystem Inflammatory Syndrome Temporally Associated with SARS-CoV-2 of the Asociación Española de Pediatría

Sylvia Belda Hofheinz (SECIP, Hospital Universitario 12 de Octubre de Madrid), Inmaculada Calvo Penadés (SERPE, Hospital Universitario y Politécnico La Fe), Juan Carlos de Carlos Vicente (SECIP, Hospital Universitario Son Espases), Carlos Daniel Grasa Lozano (SEIP, Hospital Universitario La Paz), Susanna Hernández Bou (SEUP, Hospital Sant Joan de Déu), Rosa M. Pino Ramírez (SEPHO, Hospital Sant Joan de Déu),

Esmeralda Núñez Cuadros (SERPE, Hospital Regional Universitario de Málaga), Javier Pérez-Lescure Picarzo (SECPCC, Hospital Universitario Fundación Alcorcón), Jesús Saavedra Lozano (SEIP, Hospital General Universitario Gregorio Marañón), Diana Salas-Mera (SECPCC, Hospital Universitario La Paz), Enrique Villalobos Pinto (SEPHO, Hospital Infantil Universitario Niño Jesús).

References

1. COVID-19/Informe nº 33. Análisis de los casos de COVID-19 hasta el 10 de mayo en España a 29 de mayo de 2020.pdf. 2020. Accessed 30 June 2020, at <https://www.isciii.es/QueHacemos/Servicios/VigilanciaSaludPublicaRENAVE/EnfermedadesTransmisibles/Documents/INFORMES/Informes>.
2. Estudio ENE-COVID19: segunda ronda estudio nacional de sero-epidemiología de la infección por SARS-CoV-2 en España. 2020. Accessed June 16, 2020., at https://www.mscbs.gob.es/ciudadanos/ene-covid/docs/ESTUDIO_ENE-COVID19_SEGUNDA_RONDA_INFORME_PRELIMINAR.pdf.
3. Moraleda C, Serna-Pascual M, Soriano-Arandes A, Simó S, Epalza C, Santos M, et al. Multi-Inflammatory Syndrome in Children related to SARS-CoV-2 in Spain. *Clin Infect Dis*. 2020, <http://dx.doi.org/10.1093/cid/ciaa1042>.
4. Emergency Preparedness and Response Multisystem Inflammatory Syndrome in Children (MIS-C) Associated with Coronavirus Disease; 2020. Accessed Accessed June 16, 2020., at https://emergency.cdc.gov/coca/calls/2020/callinfo_051920.asp.
5. Guidance: paediatric multisystem inflammatory syndrome temporally associated with COVID-19; 2020. Accessed 6 de junio de 2020, at <https://www.rcpch.ac.uk/resources/guidance-paediatric-multisystem-inflammatory-syndrome-temporally-associated-covid-19>.
6. Multisystem inflammatory syndrome in children and adolescents with COVID-19; 2020. Accessed June 16, 2020, at <https://www.who.int/news-room/commentaries/detail/multisystem-inflamatory-syndrome-in-children-and-adolescents-with-covid-19>.
7. Rapid risk assessment: Paediatric inflammatory multisystem syndrome and SARS-CoV-2 infection in children; 2020. Accessed 19 june 2020, at <https://www.ecdc.europa.eu/en/publications-data/paediatric-inflammatory-multisystem-syndrome-and-sars-cov-2-rapid-risk-assessment>.
8. Abdel-Mannan O, Eyre M, Lobel U, Bamford A, Eltze C, Hameed B, et al. Neurologic and Radiographic Findings Associated With COVID-19 Infection in Children. *JAMA Neurol*. 2020, <http://dx.doi.org/10.1001/jamaneurol.2020.2687>.
9. McCrindle BW, Rowley AH, Newburger JW, Burns JC, Bolger AF, Gewitz M, et al. Diagnosis, Treatment, and Long-Term Management of Kawasaki Disease: A Scientific Statement for Health Professionals From the American Heart Association. *Circulation*. 2017;135, e927-e99. doi:10.1161/CIR.0000000000000484.
10. Whittaker E, Bamford A, Kenny J, Kaforou M, Jones CE, Shah P, et al. Clinical Characteristics of 58 Children With a Pediatric Inflammatory Multisystem Syndrome Temporally Associated With SARS-CoV-2. *JAMA*. 2020, <http://dx.doi.org/10.1001/jama.2020.10369>.
11. Weiss SL, Peters MJ, Alhazzani W, Agus MSD, Flori HR, Inwald DP, et al. Surviving Sepsis Campaign International Guidelines for the Management of Septic Shock and Sepsis-Associated Organ Dysfunction in Children. *Pediatr Crit Care Med*. 2020;21, e52-e106. doi:10.1097/PCC.0000000000002198.
12. Paediatric Inflammatory Multi-system Syndrome - temporally associated with SARS-CoV 2 (PIMS-TS): Critical Care guidance; 2020. Accessed 16 de junio de 2020, at <https://pccsociety.uk/wp-content/uploads/2020/08/PIMS-TS-Critical-Care-Clinical-Guidance-v4.pdf>.
13. Belhadjer Z, Meot M, Bajolle F, Khraiche D, Legendre A, Abakka S, et al. Acute heart failure in multisystem inflammatory syndrome in children (MIS-C) in the context of global SARS-CoV-2 pandemic. *Circulation*. 2020, <http://dx.doi.org/10.1161/CIRCULATIONAHA.120.048360>.
14. Verdoni L, Mazza A, Gervasoni A, Martelli L, Ruggeri M, Ciuffreda M, et al. An outbreak of severe Kawasaki-like disease at the Italian epicentre of the SARS-CoV-2 epidemic: an observational cohort study. *Lancet*. 2020;395:1771-8, [http://dx.doi.org/10.1016/S0140-6736\(20\)31103-X](http://dx.doi.org/10.1016/S0140-6736(20)31103-X).
15. Kaushik S, Aydin SI, Derespina KR, Bansal PB, Kowalsky S, Trachtman R, et al. Multisystem Inflammatory Syndrome in Children Associated with Severe Acute Respiratory Syndrome Coronavirus 2 Infection (MIS-C): A Multi-institutional Study from New York City. *J Pediatr*. 2020;224:24-9, <http://dx.doi.org/10.1016/j.jpeds.2020.06.045>.
16. Riphagen S, Gomez X, Gonzalez-Martinez C, Wilkinson N, Theocharis P. Hyperinflammatory shock in children during COVID-19 pandemic. *Lancet*. 2020;395:1607-8, [http://dx.doi.org/10.1016/s0140-6736\(20\)31094-1](http://dx.doi.org/10.1016/s0140-6736(20)31094-1).
17. Toubiana J, Poirault C, Corsia A, Bajolle F, Fourgeaud J, Angoulvant F, et al. Kawasaki-like multisystem inflammatory syndrome in children during the covid-19 pandemic in Paris, France: prospective observational study. *BMJ*. 2020;369:m2094, <http://dx.doi.org/10.1136/bmj.m2094>.
18. Wang F, Nie J, Wang H, Zhao Q, Xiong Y, Deng L, et al. Characteristics of Peripheral Lymphocyte Subset Alteration in COVID-19 Pneumonia. *J Infect Dis*. 2020;221:1762-9, <http://dx.doi.org/10.1093/infdis/jiaa150>.
19. Bordoni V, Sacchi A, Cimini E, Notari S, Grassi G, Tartaglia E, et al. An inflammatory profile correlates with decreased frequency of cytotoxic cells in COVID-19. *Clin Infect Dis*. 2020, <http://dx.doi.org/10.1093/cid/ciaa577>.
20. Cabrero-Hernandez M, Garcia-Salido A, Leo-Gordillo I, Alonso-Cadenas JA, Gochi-Valdovinos A, Gonzalez Brabin A, et al. Severe SARS-CoV-2 Infection in Children With Suspected Acute Abdomen: A Case Series From a Tertiary Hospital in Spain. *Pediatr Infect Dis J*. 2020;39, e195-e8. doi:10.1097/INF.0000000000002777.
21. Henderson LA, Canna SW, Schulert GS, Volpi S, Lee PY, Kernan KF, et al. On the Alert for Cytokine Storm: Immunopathology in COVID-19. *Arthritis Rheumatol*. 2020;72:1059-63, <http://dx.doi.org/10.1002/art.41285>.
22. Lou B, Li TD, Zheng SF, Su YY, Li ZY, Liu W, et al. Serology characteristics of SARS-CoV-2 infection after exposure and post-symptom onset. *Eur Respir J*. 2020;56, <http://dx.doi.org/10.1183/13993003.00763-2020>.
23. Sethuraman N, Jeremiah SS, Ryo A. Interpreting Diagnostic Tests for SARS-CoV-2. *Jama*. 2020, <http://dx.doi.org/10.1001/jama.2020.8259>.
24. Tagarro A, Epalza C, Santos M, Sanz-Santaeufemia FJ, Otheo E, Moraleda C, et al. Screening and Severity of Coronavirus Disease 2019 (COVID-19) in Children in Madrid, Spain. *JAMA Pediatr*. 2020, <http://dx.doi.org/10.1001/jampediatics.2020.1346>.
25. Feldstein LR, Rose EB, Horwitz SM, Collins JP, Newhams MM, Son MBF, et al. Multisystem Inflammatory Syndrome in U.S. Children and Adolescents. *N Engl J Med*. 2020;383:334-46, <http://dx.doi.org/10.1056/NEJMoa2021680>.
26. Barrios Tascon A, Centeno Malfaz F, Rojo Sombrero H, Fernandez-Cooke E, Sanchez-Manubens J, Perez-Lescure Picarzo J, et al. [National consensus on the cardiological treatment and follow-up of Kawasaki disease]. *An Pediatr (Barc)*. 2018;89, 188 e1- e22. doi:10.1016/j.anpedi.2018.04.003.
27. Verbeek JH, Ijaz S, Mischke C, Ruotsalainen JH, Mäkelä E, Neuvonen K, et al. Personal protective equipment for preventing highly infectious diseases due to

- exposure to contaminated body fluids in healthcare staff. *Cochrane Database Syst Rev.* 2016;4:Cd011621, <http://dx.doi.org/10.1002/14651858.CD011621.pub2>.
28. Davies P, Evans C, Kanthimathinathan HK, Lillie J, Brierley J, Waters G, et al. Intensive care admissions of children with paediatric inflammatory multisystem syndrome temporally associated with SARS-CoV-2 (PIMS-TS) in the UK: a multicentre observational study. *Lancet Child Adolesc Health.* 2020;4:669–77, [http://dx.doi.org/10.1016/S2352-4642\(20\)30215-7](http://dx.doi.org/10.1016/S2352-4642(20)30215-7).
 29. Cinesi Gomez C, Penuelas Rodriguez O, Lujan Torne ML, Egea Santaolalla C, Masa Jimenez JF, Garcia Fernandez J, et al. Clinical Consensus Recommendations Regarding Non-Invasive Respiratory Support in the Adult Patient with Acute Respiratory Failure Secondary to SARS-CoV-2 infection. *Rev Esp Anestesiol Reanim.* 2020;67:261–70, <http://dx.doi.org/10.1016/j.redar.2020.03.006>.
 30. Pediatric Acute Lung Injury Consensus Conference G. Pediatric acute respiratory distress syndrome: consensus recommendations from the Pediatric Acute Lung Injury Consensus Conference. *Pediatr Crit Care Med.* 2015;16:428–39, <http://dx.doi.org/10.1097/PCC.0000000000000350>.
 31. Ramcharan T, Nolan O, Lai CY, Prabhu N, Krishnamurthy R, Richter AG, et al. Paediatric Inflammatory Multisystem Syndrome: Temporally Associated with SARS-CoV-2 (PIMS-TS): Cardiac Features, Management and Short-Term Outcomes at a UK Tertiary Paediatric Hospital. *Pediatr Cardiol.* 2020;1:1–11, <http://dx.doi.org/10.1007/s00246-020-02391-2>.
 32. Clinical Guidance for Pediatric Patients with Multisystem Inflammatory Syndrome in Children (MIS-C) Associated with SARS-CoV-2 and Hyperinflammation in COVID-19; 2020. Accessed 23 de junio de 2020, at <https://www.rheumatology.org/Portals/0/Files/ACR-COVID-19-Clinical-Guidance-Summary-MIS-C-Hyperinflammation.pdf>.
 33. Mehta P, Cron RQ, Hartwell J, Manson JJ, Tattersall RS. Silencing the cytokine storm: the use of intravenous anakinra in haemophagocytic lymphohistiocytosis or macrophage activation syndrome. *Lancet Rheumatol.* 2020;2, e358–e67. doi:10.1016/S2665-9913(20)30096-5.
 34. Guaraldi G, Meschiari M, Cozzi-Lepri A, Milic J, Tonelli R, Menozzi M, et al. Tocilizumab in patients with severe COVID-19: a retrospective cohort study. *Lancet Rheumatol.* 2020;2, e474–e84. doi:10.1016/S2665-9913(20)30173-9.
 35. Nozawa T, Imagawa T, Ito S. Coronary-Artery Aneurysm in Tocilizumab-Treated Children with Kawasaki's Disease. *N Engl J Med.* 2017;377:1894–6, <http://dx.doi.org/10.1056/NEJMc1709609>.
 36. Sarma P, Kaur H, Kumar H, Mahendru D, Avti P, Bhattacharyya A, et al. Virological and clinical cure in COVID-19 patients treated with hydroxychloroquine: A systematic review and meta-analysis. *J Med Virol.* 2020;92:776–85, <http://dx.doi.org/10.1002/jmv.25898>.
 37. Cavalcanti AB, Zampieri FG, Rosa RG, Azevedo LCP, Veiga VC, Avezum A, et al. Hydroxychloroquine with or without Azithromycin in Mild-to-Moderate Covid-19. *N Engl J Med.* 2020, <http://dx.doi.org/10.1056/NEJMoa2019014>.
 38. Rosenberg ES, Dufort EM, Udo T, Wilberschied LA, Kumar J, Tesoriero J, et al. Association of Treatment With Hydroxychloroquine or Azithromycin With In-Hospital Mortality in Patients With COVID-19 in New York State. *Jama.* 2020, <http://dx.doi.org/10.1001/jama.2020.8630>.
 39. Wang Y, Zhang D, Du G, Du R, Zhao J, Jin Y, et al. Remdesivir in adults with severe COVID-19: a randomised, double-blind, placebo-controlled, multicentre trial. *Lancet.* 2020;395:1569–78, [http://dx.doi.org/10.1016/S0140-6736\(20\)31022-9](http://dx.doi.org/10.1016/S0140-6736(20)31022-9).
 40. Shekar K, Badulak J, Peek G, Boeken U, Dalton HJ, Arora L, et al. Extracorporeal Life Support Organization COVID-19 Interim Guidelines. *ASAIO J.* 2020, <http://dx.doi.org/10.1097/MAT.0000000000001193>.